·基础研究 ·

# 新型复合人工骨的生物相容和成骨性能研究△

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摘要: [目的] 探讨脱细胞骨基质/磷酸钙/半水硫酸钙(ACBM/CPC/CSH)复合人工骨材料的生物相容性及成骨性能。[方法] 物理混合法合成 ACBM/CPC/CSH 复合人工骨材料,制备材料浸提液,用于急性、亚急性毒理、热原和表皮刺激实验。材料肌肉埋植实验,第7、14和21d取材组织病理学检查,观察肌肉炎症反应;第14d取材行流式细胞检查,观察血液与组织CD4\*、CD8\*T淋巴细胞含量变化,CCK-8法检测细胞毒性。建立大鼠股骨髁骨缺损模型,分别植入ACBM/CPC/CSH骨材料组(材料组)、磷酸钙/半水硫酸钙(CPC/CSH)(对照组)和不植任何材料(空白组),观察成骨情况。[结果]浸提液未引起小鼠急性与亚急性毒理反应,体重变化差异无统计学意义(P>0.05)。热原实验体温正常。表皮注射实验 72h内皮肤未出现红斑。肌肉埋植实验,第7d肌肉组织有轻微炎症反应,21d炎症消失;第14d流式细胞学检查,材料组、对照组和空白组外周血 CD4\* [(9.6±1.8) vs (10.1±1.2) vs (10.7±1.4), P=0.470],CD8\* [(9.5±1.1) vs (10.3±1.8) vs (10.5±1.7), P=0.249]; 脾脏 CD4\* [(18.1±1.5) vs (17.2± 7.3) vs (17.5±1.0), P=0.195],CD8\* [(8.8±7.2) vs (7.7±7.6) vs (7.8±7.2), P=0.359] 淋巴细胞含量差异无统计学意义 (P>0.05)。X线片与Miero-CT 结果显示,各组骨缺损均有不同程度修复,ACBM/CPC/CSH 骨材料修复效果最佳。[结论] ACBM/CPC/CSH 复合人工骨 材料具有良好的生物相容性和体内成骨性能,有利于不规则骨缺损的修补。

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Biocompatibility and osteogenic properties of a novel composited artificial bone // SUN Jia-pei<sup>1,2</sup>, DU Jian<sup>2</sup>, PENG Wei<sup>1</sup>. 1. Senior Department of Orthopaedics, The Fourth Medical Center of PLA General Hospital, Beijing 100142, China; 2. Hebei North University, Zhangjiakou 075132, China

Abstract: [Objective] To investigate the biocompatibility and osteogenic properties of acellular bone matrix/calcium phosphate/calcium sulfate hemihydrate (ACBM/CPC/CSH) composite artificial bone. [Methods] ACBM/CPC/CSH composite was synthesized by physical mixing method, and the material extract was prepared for acute and subacute toxicity, pyrogen and epidermal irritation tests. The material was implanted in the muscle and histopathological examination was performed on the 7th, 14th and 21d to observe the inflammatory reaction of the muscle. On the 14th day, flow cytometry was taken to observe the changes of CD4+ and CD8+T lymphocyte content in blood and tissues, and the cytotoxicity was detected by CCK-8 method. Bone defect model was established in the rat femoral condyle, and the bone formation was observed by X-ray and CT after implantation of ACBM/CPC/CSH group (material group), calcium phosphate/calcium sulfate hemihydrate (CPC/CSH) group (control group) and no material implantation (blank group). [Results] The extract did not cause acute or subacute toxicity in mice, and there was no significant difference in body weight (P>0.05). In the pyrogen test, the body temperature remained normal. Skin erythema did not appear within 72 h after epidermal injection. In the muscle implantation experiment, there was slight inflammation in the muscle tissue on the 7th day, and the inflammation disappeared on the 21st day. As results of flow cytometry on the 14th day, there were no significant differences among the three groups in terms of the peripheral blood lymphocyte content of CD4<sup>+</sup> [(9.6±1.8) vs (10.1±1.2) vs (10.7± 1.4), P=0.470], CD8<sup>+</sup> [(9.5±1.1) vs (10.3±1.8) vs (10.5±1.7), P=0.249], as well as the spleen CD4<sup>+</sup> [(18.1±1.5) vs (17.2±7.3) vs (17.5±1.0), P=0.249], as well as the spleen CD4<sup>+</sup> [(18.1±1.5) vs (17.2±7.3) vs (17.5±1.0), P=0.249], as well as the spleen CD4<sup>+</sup> [(18.1±1.5) vs (17.2±7.3) vs (17.5±1.0), P=0.249], as well as the spleen CD4<sup>+</sup> [(18.1±1.5) vs (17.2±7.3) vs (17.5±1.0), P=0.249], as well as the spleen CD4<sup>+</sup> [(18.1±1.5) vs (17.2±7.3) vs (17.5±1.0), P=0.249], as well as the spleen CD4<sup>+</sup> [(18.1±1.5) vs (17.2±7.3) vs (17.5±1.0), P=0.249], as well as the spleen CD4<sup>+</sup> [(18.1±1.5) vs (17.2±7.3) vs (17.5±1.0), P=0.249], as well as the spleen CD4<sup>+</sup> [(18.1±1.5) vs (17.2±7.3) vs (17.5±1.0), P=0.249], as well as the spleen CD4<sup>+</sup> [(18.1±1.5) vs (17.5±1.0), P=0.249], as well as the spleen CD4<sup>+</sup> [(18.1±1.5) vs (17.5±1.0), P=0.249], as well as the spleen CD4<sup>+</sup> [(18.1±1.5) vs (17.5±1.0), P=0.249], as well as the spleen CD4<sup>+</sup> [(18.1±1.5) vs (17.5±1.0), P=0.249], as well as the spleen CD4<sup>+</sup> [(18.1±1.5) vs (17.5±1.0), P=0.249], as well as the spleen CD4<sup>+</sup> [(18.1±1.5) vs (17.5±1.0), P=0.249], as well as the spleen CD4<sup>+</sup> [(18.1±1.5) vs (17.5±1.0), P=0.249], as well as the spleen CD4<sup>+</sup> [(18.1±1.5) vs (17.5±1.0), P=0.249], as well as the spleen CD4<sup>+</sup> [(18.1±1.5) vs (17.5±1.0), P=0.249], as well as the spleen CD4<sup>+</sup> [(18.1±1.5) vs (17.5±1.0), P=0.249], as well as the spleen CD4<sup>+</sup> [(18.1±1.5) vs (17.5±1.0)], as well as the spleen CD4<sup>+</sup> [(18.1±1.5) vs (17.5±1.0)], as well as the spleen CD4<sup>+</sup> [(18.1±1.5) vs (17.5±1.0)], as well as the spleen CD4<sup>+</sup> [(18.1±1.5) vs (17.5±1.0)], as well as the spleen CD4<sup>+</sup> [(18.1±1.5) vs (17.5±1.0)], as well as the spleen CD4<sup>+</sup> [(18.1±1.5) vs (17.5±1.0)], as well as the spleen CD4<sup>+</sup> [(18.1±1.5) vs (17.5±1.0)], as well as the spleen CD4<sup>+</sup> [(18.1±1.5) vs (17.5±1.0)], as well as the spleen CD4<sup>+</sup> [(18.1±1.5) vs (17.5±1.0)], as well as the spleen CD4<sup>+</sup> [(18.1±1.5) vs (17.5±1.0)], as well as the spleen CD4<sup>+</sup> [(18.1±1.5) vs (17.5±1.5)], as well as the spleen CD4<sup>+</sup> [(18.1±1.5) vs (1 0.195], and CD8<sup>+</sup> [(8.8±7.2) vs (7.7±7.6) vs (7.8±7.2), P=0.359]. The X-ray and Micro-CT images showed that bone defects in all groups were repaired to different degrees, and ACBM/CPC/CSH bone materials had the best repair effect. [Conclusion] The ACBM/CPC/CSH composite artificial bone material has good biocompatibility and in vivo osteogenic properties, which is beneficial to the repair of irregular bone defects.

Key words: acellular bone matrix, composite material, bone defect, biocompatibility, osteogenesis

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随着骨缺损疾病的不断增多,骨缺损修复材料的 制备成为目前研究热点之一。据统计,全球每年约有 两千万骨缺损患者需要接受修复治疗<sup>[1]</sup>,其中不同创 伤机制导致的各种不规则骨缺损占大多数,虽不影响 功能状态,但为追求完美基本会选择矫正整形治疗。 目前,国内外的研究通常将自体骨移植和同种异体骨 移植视为治疗骨缺损的首选方法<sup>[2,3]</sup>。然而,这种方 法仍存在局限性,例如,自体骨移植在取自体骨的同 时,必然会对身体造成一定程度的损害;同种异体骨 移植可能会诱发机体免疫排斥反应,失败率高达 10%~50%,主要表现为移植骨的延迟愈合、不完全 愈合和疲劳性骨折等<sup>[4,5]</sup>。

近期人工合成生物材料替代传统骨移植的方法, 引起了广泛的关注<sup>[6]</sup>。为了解决既不伤害自身,又能解 决不规则缺损修复与移植骨免疫排斥性的难题,笔者 开发了一种新型可塑性骨移植复合材料,以脱细胞骨 基质 (acellular bone matrix, ACBM)为核心,空隙填充 可注射性磷酸钙水泥 (calcium phosphate cement, CPC)和半水硫酸钙 (calcium sulfate hemihydrate, CSH)的脱细胞骨基质/磷酸钙/半水硫酸钙 (ACBM/ CPC/CSH)复合人工骨材料<sup>[7,8]</sup>。研究表明,ACBM有 助于细胞和生长因子的附着,并能有效降低疾病传播的 风险,本团队使用牛股骨远端干骺松质骨制备 ACBM, 其附着率可高达 62.4%<sup>[9-12]</sup>。此外,CPC 与 CSH 两种材 料具有良好的生物相容性、骨传导性和可注射性等优 点,被广泛用于骨缺损修复材料的制备中,并且两者被 认为是最有潜力的临床骨修复材料<sup>[13-15]</sup>。

本研究前期已经针对 ACBM/CPC/CSH 复合人工 骨材料的理化性能进行检测,结果表明,材料固化后 具备良好的力学性能与缺损填充度,可满足对不规则 骨缺损修补的材料要求<sup>[16]</sup>。为进一步明确性质,笔 者开展了一系列体外与体内实验,以完善 ACBM/ CPC/CSH 骨材料生物相容性和成骨性检查,为不规 则骨缺损的治疗提供新的可能性和选择。

# 1 材料和方法

## 1.1 实验动物

新西兰大白兔、SD 大鼠和 C57 小鼠由北京斯贝 福生物科技有限公司提供,所有动物实验均经中国人 民解放军总医院第八医学中心伦理委员会批准。

## 1.2 ACBM/CPC/CSH 骨材料制备

研究材料的制备过程本团队前期已经详细论述, 具体要求参见参考文献<sup>[16-19]</sup>。 1.3 体内实验

# 1.3.1 毒理实验

依据国标 GB15193.3—2014 与 GB15193.22— 2014 设计实验。(1)急性毒理实验:选取 C57 小鼠 20 只,体重 (20±2)g,雌雄参半,均分浸提液组和 生理盐水组,灌胃给药,剂量标准 10.0 g/kg。第 24、48 和 72 h 观察小鼠饮食状况并记录体重;(2) 亚急性毒理实验:分组与给药方式同上,连续灌胃 28 d,给药期间观察动物生命体征等。第 28 d 取心、 肝、脾和肾进行组织病理学检查。

1.3.2 热原实验

新西兰大白兔 5 只,体重 2.0~2.5 kg,雌雄不限,雌兔未孕。连续记录体温 3 d,选取体温变化稳定的 3 只进行实验。随机编号 1、2、3 号兔,记录正常体温,浸提液耳缘静脉注射(10 ml/kg),测定 30 min、8、24、48 和 72 h体温数值。衡量标准:72 h内,3 只兔体温升高均低于 0.6℃,且升高总和低于 1.3℃,表明该实验材料无热原物质。

1.3.3 表皮注射实验

选取3只无皮肤病新西兰大白兔, 剃除脊柱两侧 被毛, 碘伏消毒。在指定区域注射浸提液、生理盐水 和40%酒精, 观察即刻、24、48和72h各注射部位 的红斑和水肿情况。依据国标 GB/T 21604—2022标 准进行评分,并计算皮肤刺激分级。

1.3.4 炎症和免疫排斥实验

肌肉埋植实验:选取 25 只 C57 小鼠,体重 (20±2)g。炎症反应实验 10 只;免疫排斥实验 15 只,随机均分为 3 组,ACBM/CPC/CSH 骨材料组 (材料组)、同种异体骨组(对照组)和空白假手术组 (空白组)。1%戊巴比妥钠腹腔麻醉小鼠,俯卧位固 定,将材料植入大腿股后群肌袋中逐层缝合。第 7、 14 和 21 d 肌肉组织取材 HE 染色,进行组织炎症分 级。第 14 d 三组小鼠取材外周血和脾脏进行流式细 胞学检查,观察 CD4\*、CD8\*含量变化。

# 1.3.5 成骨实验

选取 SPF 级 SD 大鼠 36 只(6 周龄, 200~220 g),随机均分 3 组,每组 12 只,材料组植入 ACBM/ CPC/CSH 和对照组植入 CPC/CSH 骨材料,空白组不 填充材料。2%戊巴比妥钠(0.2 ml/100 g)腹腔麻醉 大鼠,碘伏消毒,充分暴露大鼠股骨远端,利用颅骨 钻在大鼠股骨外侧髁制备骨缺损 2 mm×3 mm。分别 植入 ACBM/CPC/CSH 骨材料(材料组)、磷酸钙/半 水硫酸钙(CPC/CSH)(对照组)和缺损中不植入任 何材料(空白组)。术后第 1、2 和 4 周 X 射线检查 评估缺损部位成骨情况,第8周股骨取材进行 Micro-CT 观察分析。

1.4 体外细胞毒性实验

制备脾淋巴细胞悬液,稀释为 2×10<sup>4</sup> 个/ml 浓 度,设立材料组(10%ACBM 浸提液+脾淋巴细胞悬 液的 RPMI1640+20%胎牛血清)、对照组(脾淋巴细 胞悬液的 RPMI1640+20%胎牛血清)和空白组(脾淋 巴细胞悬液的 RPMI1640,不加胎牛血清),每组重 复 10 孔。在 37℃、5%CO<sub>2</sub>细胞培养箱培养 72 h,加 入 CCK-8 试剂继续培养 1~4 h,多功能酶标仪上机检 测每孔吸光度。

1.5 统计学方法

采用 GraphPad.Prism.9.0 软件进行统计学分析, 计量数据用 x̄±s 表示,多组数据采用独立样本 t 检 验。P<0.05 为差异存在统计学意义。

# 2 结 果

- 2.1 体内实验
- 2.1.1 毒理实验

两组小鼠在注射后 24、48 和 72 h 体重变化差异

无统计学意义(P>0.05)(表 1),且未出现死亡昏迷、呼吸困难和四肢活动受限等现象,结果表明,该材料不会引起小鼠全身急性毒理反应。

亚急性毒理实验,两组实验小鼠生命体征良好, 未发生死亡和昏迷。实验结束对小鼠心、肝、脾和肾 进行组织病理学检查,生理盐水组箭头所示肾小球囊 壁轻微增厚(图1),其余均正常。结果表明该材料 不会引起小鼠亚急性毒理实验。

表 1. 两组急性毒理实验小鼠体重(g, x̄±s)与比较							
Table 1. Body weight of mice in acute toxicology experiments							
$(g, \ \overline{x} \pm s)$							
时间 (h)	浸提液组	生理盐水组	D店				
	( <i>n</i> =20)	( <i>n</i> =20)	Г Ц.				
0	20.2±0.3	20.2±0.5	0.695				
24	20.9±0.3	21.1±0.5	0.163				
48	21.9±0.3	22.1±0.4	0.478				
72	22.8±0.4	22.9±0.5	0.603				

2.1.2 热原实验

浸提液耳缘静脉注射,兔体温升高均低于 0.6℃,且升高总和低于 1.3℃,见表 2,热原反应阴 性。



图 1. 亚急性毒理试验组织学观察(HE, ×200, 标尺=100 μm)。1a~1d: 浸提液组; 1e~1h: 生理盐水组; 两组 HE 染色均未见 明显病理改变, 肾脏肾小球囊壁稍增厚。

Figure 1. Histological observation of subacute toxicological test (HE,  $\times 200$ , scale=100  $\mu$ m). 1a~1d: The extract group; 1e~1h: The normal saline group; HE staining showed no significant pathological changes found in both groups, and a slight thickening of the glomerular capsule wall of the kidney was seen.

## 2.1.3 表皮注射实验

注射浸提液与生理盐水处皮肤均未出现红斑 或水肿,原发性刺激指数评分约 0.3,均<0.5,刺 激强度为无刺激性(表 3),注射部位表现(图 2)。

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表 2. 热原试验兔体温 Table 2. Body temperature of pyrogen test rabbits								
动物编号 正常体温 (℃)	正常体温	注射后 (h)					升温	升温总和
	$(\mathfrak{D})$	0.5	8	24	48	72	$(\mathcal{C})$	$(\mathcal{C})$
1	39.2	39.5	39.6	39.2	39.0	39.3	0.4	0.5
2	39.0	39.0	39.9	39.0	38.7	38.8	0	0.5
3	39.4	39.2	39.5	39.4	38.9	39.1	0.1	0.5

注:浸提液注射后体温下降,升温记为0℃。升温低于0.6℃,且升温总和低于1.3℃,表明该材料检测合格。

Note: The body temperature dropped after injection of the extract solution, and the temperature rise was recorded as  $0^{\circ}$ C. The temperature rise is lower than  $0.6^{\circ}$ C, and the total temperature rise is lower than  $1.3^{\circ}$ C, indicating that the material passes the test.

表 3. 表皮注射实验刺激评分 Table 3. Stimulation score of epidermal injection experiment						
材料	the theory with	原性刺激记分				百世世却地长来
		即刻	24 h	48 h	72 h	原反性刺激指数
实验材料浸提液	3	0	0	0	0.3	0.3
生理盐水	3	0	0	0.3	0.3	0.3
40%酒精	3	0	0.3	0.2	2.7	2.7

注: <0.5 无刺激性; 0.5~<2.0 轻刺激; 2.0~<6.0 中等刺激; 6.0~8.0 强刺激。

Note: <0.5 is non-irritating; 0.5~<2.0 is lightly irritating; 2.0~<6.0 is moderately irritating; 6.0~8.0 is strongly irritating.



图 2. 表皮注射实验兔皮肤变化。

Figure 2. Skin changes of rabbits in epidermal injection test.

## 2.1.4 炎症和免疫排异实验

炎症分级:肉眼观小鼠手术处皮肤无红肿变化, 第7d病理学观察有轻微炎性反应,第21d未见炎性 细胞浸润,炎性反应彻底消失(图3)。

免疫排异实验:第14d取材外周血和脾脏样本,进行流式细胞学检查,结果见表4,材料组与另外两组外周血和组织的CD4\*和CD8\*T淋巴细胞含量差异无统计学意义(P>0.05),该实验材料不会引起免疫排异反应。

## 2.1.5 成骨实验

分别植入 ACBM/CPC/CSH 骨材料(材料组)、磷酸钙/半水硫酸钙(CPC/CSH)(对照组)和缺损中不植入任何材料(空白组)。X 射线观察见图 4。材料组第1周可见骨缺损和骨移植材料,第2、4周材料中心连续骨痂出现,缺损修补面模糊,边缘锐利性下降,第4

周更加明显,骨髓腔通畅,边缘骨密度升高,骨化程 度进一步增强。而对照组的空白组成骨差。

第8周进行 Micro-CT 整体、轴位和冠状位检查。结果显示,材料组未见骨缺损轮廓,缺损基本愈合,仅在冠状位可见少量疑似残留材料的密度增高影,骨小梁呈向心性生长排列;对照组和空白组三种观察面缺损区域较明显,影像观察可见缺损凹陷。

2.2 体外细胞毒性实验

浸提液对小鼠淋巴细胞增殖无明显抑制,细胞生长情况良好。材料组和对照组之间吸光度(OD值) [(0.170±0.008) vs (0.168±0.008), P=0.646] 差异无统计 学意义,但是,材料组 OD 值显著大于空白组 [(0.170±0.008) vs (0.134±0.004), P<0.001]。

3 讨 论

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本团队前期的研究以 ACI	BM 为主体,可注射性	好孔隙率和降解性能的 C	PC/CSH 复合材料 <sup>[19]</sup> 。CPC
CPC/CSH 复合材料以注射填充	5的形式,成功制备了	是通过酸性和碱性磷酸钙	水合反应制得,其本身已经
一种 ACBM/CPC/CSH 新型复合	·人工骨材料。这种复	具有出色的生物相容性利	II较好的成骨诱导性 <sup>[20, 21]</sup> ,

合材料具有较高的可塑性,适用于不规则骨缺损的修 复,提高缺损填充度,且具备出色的理化性能<sup>[16]</sup>。 为进一步探讨 ACBM/CPC/CSH 复合材料的生物相容 性和成骨性能,本团队进行了一系列实验。其中 CPC、CSH 复合材料本身的比例复合,形成了具有良 好北原率和降解性能的 CPC/CSH 复合材料<sup>(19)</sup>。CPC 是通过酸性和碱性磷酸钙水合反应制得,其本身已经 具有出色的生物相容性和较好的成骨诱导性<sup>[20, 21]</sup>, 同时利用 CPC/CSH 的可注射特性来填充制备材料本 身,固化反应产生的热量相对较低,固化放热性能优 于临床传统意义上的聚甲基丙烯酸甲酯(polymethyl methacrylate, PMMA)骨水泥,对周边组织损伤较 小<sup>[22]</sup>,这一材料创新为研究的临床应用提供可行性。



图 3. 炎症分级(HE, ×200, 标尺 100 µm)。3a: 植入后 7 d, 圆圈内见少量肌纤维断裂,炎症分级 2 级; 3b, 3c: 植入后第 14、21 d, 肌纤维分布均匀,炎症分级 0 级。

Figure 3. inflammatory classification (HE,  $\times 200$ , scale bar=100  $\mu$ m). 3a: Seven days after implantation small number of broken muscle fibers were seen within the the circles area, and was marked as grade 2 inflammation; 3b, 3c: Fourteen and 21 days, the muscle fibers were evenly distributed, and were marked as grade 0 inflammation.

表 4. 免疫排异实验 CD4 <sup>+</sup> 、CD8 <sup>+</sup> 含量变化 ( $\bar{x} \pm s$ , $n=5$ , %)							
	Table 4. Changes in CD4 <sup>+</sup> and CD8 <sup>+</sup> contents in immune rejection experiments ( $\bar{x} \pm s, n=5, \%$ )						
组别	材料组 (n=5)	对照组 (n=5)	空白组 (n=5)	<i>P</i> 值			
外周血 CD4+	9.6±1.8	10.1±1.2	10.7±1.4	0.470			
外周血 CD8+	9.5±1.1	10.3±1.8	10.5±1.7	0.249			
脾脏 CD4+	18.1±1.5	17.2±7.3	17.5±1.0	0.195			
脾脏 CD8+	8.8±7.2	7.7±7.6	7.8±7.2	0.359			

生物相容性作为生物材料研究制备的核心主题, 本团队深入探究了 ACBM/CPC/CSH 复合人工材料的 生物安全性、免疫排斥性和细胞毒性<sup>[23]</sup>。体内外实 验结果显示,研究材料植入动物体内,与生物组织环 境相互影响,最终能够达到平衡状态直到植入材料被 降解吸收,这一现象表明 ACBM/CPC/CSH 复合人工 骨材料具有优异的生物相容性,与之前的研究相符。 Hu 等<sup>[19]</sup> 成功引入硫酸钙 (calcium sulfate, CS) 到 CPC 中,形成硫酸钙/磷酸钙骨水泥 (calcium sulphate/phosphate cement, CSPC) 骨材料,材料本身在 动物实验中同样展现出出色的生物相容性、降解性和 成骨诱导能力。为进一步评估材料的成骨性能,本研 究制备了大鼠股骨外侧髁骨缺损模型,并在第1、 2、4 周利用 X 射线和第8 周 Micro-CT 进行骨缺损骨 化进展观察,结果显示,第1、2周,各组骨缺损骨 化进展基本相似, 第4周材料组骨化程度明显优于其 余两组,第8周材料组缺损区域已完全骨化,骨髓腔 通畅,股骨外侧髁缺损骨密度增高。结果表明,相较 单一 CPC/CSH 材料,添加了 ACBM 的复合人工骨, 成骨性能最佳,其 ACBM 自身的天然结构,如骨小 梁等,为成骨细胞的迁移和增殖提供了天然支架,材 料的细胞黏附性强<sup>[9-12]</sup>,从而导致了其卓越的成骨性 能。与之前研究成果一致,Huang等<sup>[24]</sup>成功制备的 一种基于 ACBM 的癌细胞归巢肿瘤加速器,孙晓雷 等<sup>[11]</sup>对牛松质骨 ACBM 进行的一系列研究,结果均 表明 ACBM 本身具有较好的生物力学性能、孔隙率 及黏附性能,有助于加速成骨细胞的生长增殖。结果 表明,ACBM/CPC/CSH 复合人工骨材料不仅具有良 好的生物相容性,同时具备优越的成骨性能。

综上所述,基于 ACBM 制备的 ACBM/CPC/CSH 复合人工骨材料具有出色的可塑性和缺损填充度,适 用于不规则骨缺损处的修复,此研究材料的可塑性不 仅能够迅速适应各种不规则形状骨缺损,同时其卓越 的生物相容性和成骨性能进一步完善了研究材料,提 高其在临床转化的可能性。为进一步缩短材料成骨周 期,可以考虑在这一基础上引入相关转染细胞,向组 织工程骨研究领域转化。本研究为多种创伤导致的不 规则小块状骨缺损的治疗提供了更好的选择,并且有 望在未来的临床实践中发挥重要作用。



图 4. 股骨髁骨缺损植入试验影像观察。纵向排列从上至下分别是 1、2、4 周大鼠股骨侧位 X 线片, 8 周大鼠股骨三维 Micro-CT 重建,以及横切面和冠状面 Micro-CT 片。横向排列从左向右依次为植入 ACBM/CPC/CSH 骨材料(材料组)、CPC/ CSH (对照组),和缺损中不植入任何材料(空白组)。可见材料组成骨明显优于对照组和空白组。

Figure 4. Imaging observation of osteogenesis in femoral condylar bone defect for implantation test. The longitudinal arrangement from top to bottom was the lateral X-ray of the rat femur at 1, 2, and 4 weeks, as well as the three-dimensional Micro-CT reconstruction, the transverse and coronal Micro-CT images respectively at 8 weeks. Transverse arrangement from left to right was of implantation of ACBM/ CPC/CSH artificial bone (the material group), CPC/CSH (the control group), and no material implanted in the defect (blank group). It can be seen that bone formation in the material group is significantly better than that in the control group and the blank group.

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